Disease Note

FIRST REPORT OF *CUCUMBER MOSAIC VIRUS* AND *OLIVE LATENT RINGSPOT VIRUS* ON OLIVE TREES IN LEBANON

E. Choueiri[1], A. Freiji[2], R. Abou Kubaa[2] and T. Elbeaino[2]

[1]Department of Plant Protection, Lebanese Agricultural Research Institute, Tal Amara, P.O. Box 287, Zahlé, Lebanon
[2]Istituto Agronomico Mediterraneo di Bari, Via Ceglie 9, 70010 Valenzano (Bari), Italy

Olive (*Olea europea L.*) is cultivated in Lebanon on a surface of *ca.* 53.600 ha, accounting for 43% of the area given over to perennial crops (Anonymous, 2010). Olive trees are affected by a number of virus and virus-like agents that persist in propagating material, with which they can be transmitted and disseminated (Martelli, 2011). Twelve trees of cv. Airouni, which is one of the most important cultivars grown in the country, were sampled in northern Lebanon and assayed for determining their sanitary status, in view of their possible use by nurseries for propagation. No symptoms of putative viral origin were observed in the surveyed orchards. Leaf samples were tested by RT-PCR to assess the presence of viruses commonly found in olive, i.e. *Arabis mosaic virus* (ArMV), *Cherry leaf roll virus* (CLRV), *Cucumber mosaic virus* (CMV), *Olive latent ringspot virus* (OLRSV), *Olive latent virus 1* (OLV-1), *Olive latent virus 2* (OLV-2), Olive leaf yellowing-associated virus (OLYaV) and *Strawberry latent ringspot virus* (SLRSV), using the protocol and the sets of specific primers reported by Grieco *et al.* (2000). Results showed that CLRV, OLV-1, OLV-2 and OLYaV are present in nine trees, whereas ArMV and SLRV were not found. Two samples were positive for CMV and OLRSV, yielding the expected amplified products of 513 bp and 492 bp, respectively. Both these viruses were mechanically transmitted to herbaceous hosts (*Cucumis sativus* and *Chenopodium quinoa*, respectively) and their presence in these hosts was ascertained by RT-PCR. One of the trees proved free from all viruses and may constitute a source of virus-tested material for propagation. To our knowledge, this is the first report of CMV and OLRSV in olive in Lebanon.

Anonymous, 2010. Résultats globaux du module de base du recensement de l'agriculture 2010. Projet Observatoire Libanais de Développement Agricole, Beirut, Lebanon.

Grieco F., Alkowni R., Saponari M., Savino V., Martelli G.P. 2000. Molecular detection of olive viruses. *Bulletin OEPP/EPPO Bulletin* **30**: 469-473.

Martelli G.P., 2011. Infectious diseases of olive. In: Schena L., Agosteo G.E. and Cacciola S.O. (eds). Olive Diseases and Disorders, pp. 71-88. Tansworld Research Network, Trivandrum, India.

Corresponding author: E. Choueiri
Fax: +961.8.900077
E-mail: echoueiri@lari.gov.lb

Received January 12, 2015
Accepted January 30, 2015

Disease Note

FIRST REPORT OF *ITERSONILIA PERPLEXANS* ON *ANETHUM GRAVEOLENS* IN ITALY

G. Bubici

Istituto per la Protezione Sostenibile delle Piante, Sezione di Virologia, Consiglio Nazionale delle Ricerche, via Amendola 165/A, 70126 Bari, Italy

In Apulia (southern Italy), dill (*Anethum graveolens*) is cultivated in a few greenhouses and fields, mainly located between Bari and Brindisi. In December-January of 2010 to 2014, leaf blight was observed in greenhouse-grown dill crops, when they were near the harvest stage. Symptoms differed from those induced by some known foliar pathogens of dill, i.e. *Sclerotinia sclerotiorum* and *Erysiphe heraclei*, but recalled those caused by *Botrytis cinerea*. In fact, leaves that were first discolored, turned light to dark brown, and finally wilted, rendering the affected plants unsuitable for harvest. In the presence of high relative humidity, a whitish-cream mycelium developed on symptomatic leaves. Light microscope observations revealed the presence of hyaline hyphae with clamp connections and lunate ballistospores [16.4 ± 1.7 (standard deviation) × 11.0 ± 1.3 µm]. A 609 bp product amplified by PCR from fungal DNA using the ITS1/ITS4 primer pair was sequenced (BMR Genomics, Italy) and the sequence of an isolate designated IPSP-GB520 was deposited in GenBank under the accession No. KP890654. BLAST alignment revealed a 99% homology at the nucleotide level with several accessions of *Itersonilia perplexans* Derx (Basidiomycota, Cystofilobasidiaceae), including two isolates (DQ667163 and NR_077117) conserved at the CBS-KNAW Fungal Biodiversity Centre, Utrecht, The Netherlands. Koch's postulates were fulfilled with a successful detached-leaf assay. Based on morphological characteristics and nucleotide sequence homology, the fungus was identified as *I. perplexans*. In Italy, an early report ascribed leaf blight of dill to *I. pastinacae*, based on morphometric observations and pathogenicity tests (Matta and Garibaldi, 1968). To the best of my knowledge, this is the first report of *I. perplexans* on dill in Italy.

Matta A., Garibaldi A., 1968. L'*Itersonilia pastinacae* Channon su aneto. *Phytopathologia Mediterranea* **7**: 34-39.

Corresponding author: G. Bubici
Fax: +39 080 544 3608
E-mail: giovanninicola.bubici@cnr.it

Received March 9, 2015
Accepted March 10, 2015