

DISEASE NOTE

FIRST REPORT OF TWO DISTINCT STRAINS OF *PEPINO MOSAIC VIRUS* INFECTING TOMATOES IN GREENHOUSES IN LITHUANIA

D. Šneideris, M. Žižyte, I. Zitikaite,
L. Urbanavičienė and J. Staniulis

Institute of Botany Nature Research Centre, Zaliuju Ezeru 49,
08406 Vilnius, Lithuania

During 2010 and 2011, one hundred thirty one symptomatic samples of tomato leaves and fruits were collected from commercial greenhouses in Lithuania and analyzed for the presence of *Pepino mosaic virus* (PepMV). Two isolates of this virus (NV and KK) were found in two different greenhouses on a tomato plant exhibiting mild yellow leaf spotting and on tomato fruits with marbling symptoms, respectively. The presence of the virus was confirmed by RT-PCR and DAS-ELISA, using commercial antibodies (DSMZ, Germany). Filamentous virus particles were observed by electron microscopy. The coat protein (CP) gene of the two viral isolates was amplified using a two-step RT-PCR with the PepTGB-F/PepUTR-R primer pair (Mumford and Metcalfe, 2001). A specific 845 bp PCR product was obtained, purified and sequenced. The nucleotide sequence of the CP gene of PepMV-NV and -KK was submitted to GenBank under the accession numbers JQ979169 and JQ979170, respectively. Nucleotide sequence analysis showed that the two Lithuanian PepMV isolates differ from each other and belong to two distinct PepMV genotypes. PepMV-NV belongs to the EU genotype while PepMV-KK was assigned to the CH2 genotype. These results are consistent with an expansion of the distribution of PepMV in many European countries, including Poland (Pospieszny *et al.*, 2008). They also show that both PepMV EU and CH2 genotypes are becoming common in Europe.

Mumford R.A., Metcalfe E.J., 2001. The partial sequencing of the genomic RNA of a UK isolate of *Pepino mosaic virus* and the comparison of the coat protein sequence with other isolates from Europe and Peru. *Archives of Virology* **146**: 2455-2460.

Pospieszny H., Hásiow B., Borodynyko N., 2008. Characterization of two distinct Polish isolates of *Pepino mosaic virus*. *European Journal of Plant Pathology* **122**: 443-445.

Corresponding author: D. Šneideris
Fax: +370 5 2729950
E-mail: donatas.sneideris@gmail.com

Received September 3, 2012
Accepted November 15, 2012

DISEASE NOTE

FIRST REPORT OF *GIBBERELLA AVENACEA* CAUSING WET APPLE CORE ROT IN ITALY

S.M. Sanzani¹, C. Cariddi¹, A. Roccotelli²,
F. Garganese¹, F. Fallanaj¹ and A. Ippolito¹

¹Dipartimento di Scienze del Suolo, della Pianta e degli Alimenti,
Università degli Studi Aldo Moro,
Via Amendola 165/A, 70126 Bari, Italy

²Dipartimento di Gestione dei Sistemi Agrari e Forestali,
Università Mediterranea,
Località Feo di Vito, 89122 Reggio Calabria, Italy

Gibberella avenacea R.J. Cook, [syn. *Fusarium avenaceum* (Fr.) Sacc.] is a widely distributed plant pathogen and the causal agent of wet apple core rot (wACR), a disease that develops inside the fruits and remains undetected until they are eaten (Sørensen *et al.*, 2009). In 2012, wACR symptoms were observed in apples cv. Golden Delicious from a southern Italian orchard. Diseased carpel tissues were colonized by a white mycelium, whereas the surrounding mesocarp tissues showed a light-brown wet rot. Single spore cultures were grown on PDA slants and carnation leaf agar (CLA) plates. On PDA slants, after 14 days at alternating day/night temperatures of 25/20°C and a 12 h photoperiod, the fungus formed abundant floccose white mycelium with pale orange sporodochia and released a greyish-rose pigment in the agar. Macroconidia formed on CLA were 40-80×3.5-4 µm, slightly falcate, thin-walled, usually 5 septate, with a tapering apical cell. Microconidia and chlamydospores were absent. Based on these morphological characters the fungus was identified as *G. avenacea*. For molecular confirmation, DNA was extracted from the fungal mycelium (Sanzani *et al.*, 2012), its internal transcribed spacer regions ITS1 and ITS2, including the 5.8S gene, were amplified using the universal primers ITS5/ITS4 (White *et al.*, 1990) and sequenced (GenBank KC342826). BLAST analysis of the 424 bp amplicon showed 100% identity with other *G. avenacea*/*F. avenaceum* ITS sequences from database. This is the first report of *G. avenacea* as causal agent of wACR in Italy. *G. avenacea* infections constitute an economical problem for growers and a safety issue due to the potential production of mycotoxins such as moniliformin and enniatins (Sørensen *et al.*, 2009).

Sanzani S.M., Schena L., De Cicco V., Ippolito A., 2012. Detection and quantification of *Botrytis cinerea* in symptomless table grape stamens and berries. *Postharvest Biology and Technology* **68**: 64-71.

Sørensen J.L., Phipps R.K., Nielsen K.F., Schroers H.-J., Frank J., Thrane U., 2009. Analysis of *Fusarium avenaceum* metabolites produced during wet apple core rot. *Journal of Agriculture and Food Chemistry* **57**: 1632-1639.

White T.J., Bruns T., Lee S., Taylor J.W., 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M.A., Gelfand D.H., Sninsky J.J., White T.J. (eds). *PCR Protocols: A Guide to Methods and Applications*, pp. 315-322. Academic Press, New York, NY, USA.

Corresponding author: S.M. Sanzani
Fax: +39.080.5442911
E-mail: simonamarianna.sanzani@uniba.it

Received December 20, 2012
Accepted January 2nd, 2013

DISEASE NOTE

**FIRST REPORT OF *GRAPEVINE
RUPESTRIS STEM PITTING-ASSOCIATED
VIRUS* IN TUNISIAN GRAPEVINES**

**I. Soltani¹, N. Mahfoudhi¹, T. Elbeaino², M. Digiario²
and M.R. Hajlaoui¹**

¹Laboratoire de Protection des Végétaux, Institut National
de la Recherche Agronomique de Tunisie,
Rue Hédi Karray, 2049 Ariana, Tunisia

²Istituto Agronomico Mediterraneo, Via Ceglie 9,
70010 Valenzano, Bari, Italy

Grapevine rupestris stem pitting-associated virus (GRSPaV), a member of the genus *Foveavirus*, is associated with Rupestris stem pitting, a disease that, along with Kober stem grooving (KSG), Corky bark (CB) and LN33 stem grooving, constitute the rugose wood disease complex of grapevine. *Grapevine virus A* (GVA) and *Grapevine virus B* (GVB), which are associated with the KSG and CB, respectively (Martelli, 1993), are common in Tunisian grapevines (Mahfoudhi *et al.*, 1998), whereas no information is available on the presence of GRSPaV. Therefore, 140 samples of 10 table grape varieties (Italia, Superior Seedless, Down Seedless, Red Globe, Victoria, Early Sugar, Black Pearl, King's Ruby, Rich Baba and Sultanine) from different Tunisian grapevine-growing areas were collected and indexed for the presence of this virus by RT-PCR using as template total nucleic acid preparations extracted from phloem tissue (Foissac *et al.*, 2001) and primers RSP-48 (5'-AGCTGGGATTATAAGGGAGGT-3') and RSP-49 (5'-CCAGCCGTTCCACCACTAAT-3') (Lima *et al.*, 2006). A 330 bp product corresponding to a fragment of the GRSPaV coat protein gene was amplified from 112 samples, accounting for an infection rate of 80%. The virus was present in all the surveyed areas and grape cultivars tested. More specifically, it was detected in all samples of cv. Early Sugar, King's Ruby and Sultanine, whereas in the other cultivars the infection rate ranged from 17% (cv. Down Seedless) to 97% (cv. Italia). To our knowledge, this is the first report of GRSPaV in grapevines in Tunisia.

Foissac X., Svanella -Dumas L., Gentil P., Dulucq M.J., Candresse T., 2001. Polyvalent detection of fruit tree Tricho, Capillo and Foveaviruses by nested RT-PCR using degenerated and inosine containing primers (DOP RT-PCR). *Acta Horticulturae* **550**: 37-43.

Lima M.F., Alkowni R., Uyemoto J.K., Golino D., Osman F., Rowhani A., 2006. Molecular analysis of a California strain of Rupestris stem pitting-associated virus isolated from declining Syrah grapevines. *Archives of Virology* **151**: 1889-1894.

Mahfoudhi N., Digiario M., Savino V., Di Terlizzi B., 1998. Viruses and virus diseases of grapevine in Tunisia. *Bulletin OEPP/EPPO Bulletin* **28**: 197-204.

Martelli G.P., 1993. Graft-transmissible Diseases of Grapevines. Handbook for Detection and Diagnosis. FAO Publication Division, Rome, Italy.

Corresponding author: N. Mahfoudhi
Fax: +216.71.752.897
E-mail: nmahfoudhi@yahoo.fr

Received July 19, 2010
Accepted September 24, 2010

DISEASE NOTE

**FIRST REPORT OF *HOP STUNT VIROID*
IN LEBANESE FIG TREES**

T. Elbeaino¹, E. Choueiri² and M. Digiario¹

¹Istituto Agronomico Mediterraneo, Via Ceglie 9,
70010 Valenzano (Bari), Italy

²Department of Plant Protection, Lebanese
Agricultural Research Institute, Tal Amara, P.O. Box 287,
Zablé, Lebanon

In the course of a survey carried out in 2010 in the main fig-growing areas of Lebanon (Bekaa and Mount Lebanon), a total of 100 samples representative of 15 different cultivars were collected and tested by RT-PCR for the presence of *Hop stunt viroid* (HSVd) using viroid-specific primers (Sano *et al.*, 2001). Distinct PCR products of the expected size (303 bp) were amplified from 12 fig trees of cvs Houmairi (6), Bayadi (5) and Sweidi (1), all originating from the Bekaa valley. This result was further confirmed by dot blot molecular hybridization using a HSVd-specific riboprobe. Only seven infected plants showed symptoms resembling those typical of Fig mosaic disease, the remaining being symptomless. Five of these HSVd isolates were sequenced (accession Nos HE662802-HE662806) and compared with the other HSVd isolates from GenBank. In Blast analysis, all the sequenced HSVd isolates showed 99-100% nucleotide identity among them and 94-95% identity with the HSVd type member (X0009). The only exception was the isolate D1, which confirmed the 94% identity with HSVd type but showed a lower identity level (94-95%) with the other HSVd Lebanese fig isolates. In a phylogenetic tree constructed with whole viroidal sequences the Lebanese fig isolates, except for D1, clustered in a separate group together with fig isolates from Syria and mulberry isolates from Lebanon (Elbeaino *et al.*, 2012a, 2012b). To our knowledge this is the first report of HSVd in *Ficus carica* in Lebanon.

Elbeaino T., Abou Kubaa R., Ismaeil F., Mando J., Digiario M., 2012a. Viruses and viroids of fig trees in Syria. *Journal of Plant Pathology* **94**: 687-691

Elbeaino T., Abou Kubaa R., Choueiri E., Digiario M., Navarro B., 2012b. Occurrence of Hop stunt viroid in Mulberry (*Morus alba*) in Lebanon and Italy. *Journal of Phytopathology* **160**: 148-151.

Sano T., Mimura R., Ohshima K., 2001. Phylogenetic analysis of hop and grapevine isolates of Hop stunt viroid supports a grapevine origin for hop stunt disease. *Virus Genes* **22**: 53-59.

Corresponding author: T. Elbeaino
Fax: +39.080.4606503
E-mail: elbeaino@iamb.it

Received January 2nd, 2012
Accepted January 24, 2012