

## DISEASE NOTE

**FIRST REPORT OF *LITTLE CHERRY VIRUS 2* ON *PRUNUS CERASUS* var. *MARASCA* IN CROATIA**

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*Little cherry virus 2* (LChV-2) is considered an important pathogen of cherries and, as such, testing of plant material is required according to the EPPPO certification scheme PM 4/29 for cherries. In Croatia sour cherry Marasca is a well-known native variety with total production of 1500-1800 tons of fresh fruit per year, used mainly for production of renowned liqueurs and juices. In June 2014, 19 trees were selected and total RNA was extracted from leaves with the RNeasy plant mini kit (Qiagen, Germany). RT-PCR for LChV-2 was performed using two sets of primers: LCHV-2LO2/LCHV2UP2 (Rott and Jelkmann, 2001), amplifying a 438 bp fragment of the methyltransferase (MT) gene, and LC26L/LC26R (Eastwell and Bernardy, 2001), amplifying a 409 bp fragment of the RdRp gene. Six trees were positive, four with the first primer set and two with the second. However, none of the trees tested positive by both sets, suggesting the presence of significant sequence variability among LChV-2 isolates (Theilmann *et al.*, 2004) and the presence of at least two virus variants in Croatia. One PCR product for each primer set was sequenced from both directions and sequences were compiled using MEGA6 software. BLAST searches indicated that the part of the MT gene of Croatian isolate M-79 (GenBank accession No. KT369315) shares 88% identity with the USA6b isolate of LChV-2 (AF531505), while isolate M-75 (KT369316) was closest to Canadian strain LC5 (AF416335), sharing 96% identity with part of the RdRp gene. During the 2015 harvest period, some of the LChV-2-infected trees displayed the characteristic symptoms of uneven ripening. To the best of our knowledge this is the first report of LChV-2 in Croatia.

Eastwell K.C., Bernardy M.G., 2001. Partial characterization of a closterovirus associated with apple mealybug-transmitted little cherry disease in North America. *Phytopathology* **91**: 268-273.

Rott M.E., Jelkmann W., 2001. Detection and partial characterization of a second closterovirus associated with little cherry disease, Little cherry virus 2. *Phytopathology* **91**: 261-267.

Theilmann J., Orban S., Rochon D., 2004. High sequence variability among Little cherry virus isolates occurring in British Columbia. *Plant Disease* **88**: 1092-1098.

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**FIRST MOLECULAR IDENTIFICATION OF '*CANDIDATUS PHYTOPLASMA TRIFOLIIF*' (16SrVI-D) IN *CROTON BONPLANDIANUM* FROM INDIA**

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*Croton bonplandianum* Baill., a member of the family Euphorbiaceae, is a problematic weed throughout India. During 2013, phytoplasma like symptoms of leaf yellows, little leaf and witches' broom were observed in *C. bonplandianum* plants in the fields of the Sugarcane Research Station, Gorakhpur district of Uttar Pradesh (India). Leaves were collected from four symptomatic and three symptomless plants and DNA was extracted. Fragments of ca. 1.5 kbp of 16S rDNA were amplified from all four symptomatic samples in PCR assays using the phytoplasma specific primer pair P1/P6 (Deng and Hiruki, 1991) and the amplified product was sequenced (GenBank accession No. KJ410527). No amplification was achieved from any of the symptomless leaf samples. A pairwise 16S rDNA sequence comparison revealed the highest identity (99%) of *C. bonplandianum* phytoplasma strain with a member (JX104336) of the 16SrVI group ('*Candidatus* Phytoplasma trifolii'). The phylogenetic analysis of the 16S rDNA sequence of the *Croton* phytoplasma strain also revealed its close relationship with strains of '*Ca. Phytoplasma trifolii*'. The ca. 1.5k bp product of 16S rRNA sequence of *C. bonplandianum* phytoplasma strain submitted to *iPhyClassifier* online tool assigned it to the 16SrVI-D subgroup. Earlier, Tiwari *et al.* (2014) had reported the natural occurrence of '*Ca. P. asteris*' strain in ornamental croton (*Codiaeum variegatum*) in India, whereas Naik *et al.* (2015) reported a phytoplasma association with yellows and little leaf disease of *C. bonplandianum* in India on the basis of nested PCR amplifications but no group assignment was made. Hence, the present identification of 16SrVI-D subgroup phytoplasma associated with *C. bonplandianum* constitutes the first report worldwide.

Deng S., Hiruki C., 1991. Amplification of 16S rRNA genes from culturable and nonculturable mollicutes. *Journal of Microbiological Methods* **14**: 53-61.

Naik V.K.D., Devi R.S.J., Reddy B.V.B., Shreef S.M., Rani A.R., 2015. Phytoplasma disease associated with *Croton bonplandianum* weed in Andhra Pradesh, India. *Journal of Plant Development Science* **7**: 469-470.

Tiwari A.K., Shukla K., Kumar S., Madhupriya, Rao G.P., 2014. '*Candidatus* Phytoplasma asteris' association with leaf yellows and witches' broom symptoms of *Codiaeum variegatum* in India. *Journal of Plant Pathology* **96**: S4.113-S4.131.

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